

Research article **HEALTH**

Regenerating Intervertebral Discs with Chitosan **Hydrogels**



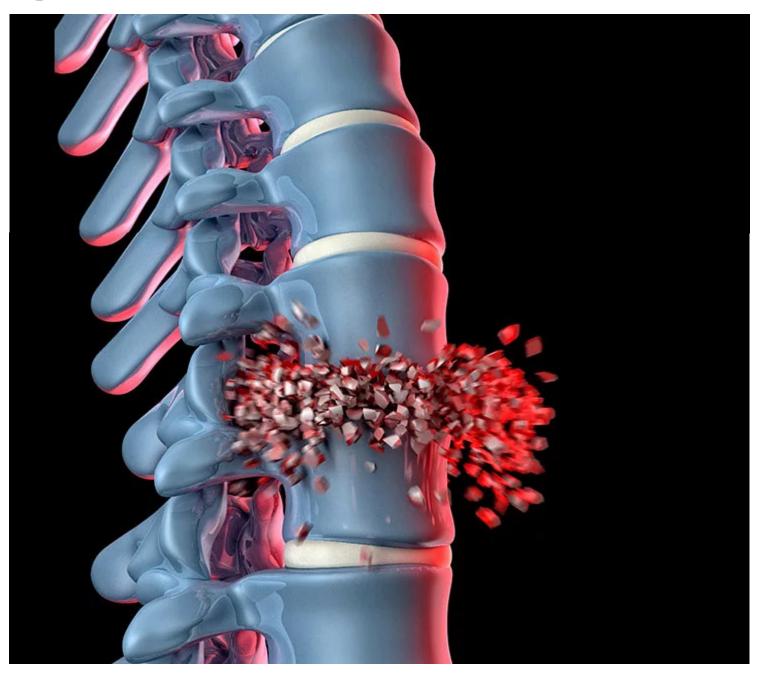












SUMMARY

Cell therapy using cells encapsulated in injectable matrices is a promising technique for the minimally invasive treatment of intervertebral disc degeneration, one of the major causes of back pain which affects 19% of the world's population. However, this therapeutic approach is limited by the generally low mechanical properties of injectable matrices. This study demonstrates the potential of new injectable chitosan hydrogels formulations with interesting mechanical properties for the regeneration of intervertebral discs, in particular their central part called nucleus pulposus (NP).

Introduction

Intervertebral disc (IVD) degeneration is one of the leading causes of lower back pain [1], which affects 19% of the worldwild population [2]. Current techniques in managing this dysfunction are not only very invasive, but also ineffective for the complete restoration of IVD biomechanical functions. Therefore, developing new minimally invasive techniques, focused on the repair or restoration of IVD, is relevant. Cell therapy is one of the promising techniques for this purpose. This technique involves injecting specific cells and molecules, potentially encapsulated in biocompatible matrices, with the objective of regenerating the disc, specifically its central part: the nucleus pulposus (NP). However, this application is limited by the generally poor mechanical properties of the injectable matrices studied so far [3, 4].

Our team has recently developed injectable chitosan hydrogels with enhanced mechanical properties and better cytocompatibility [5]. Those formulations, which are liquid at room temperature, gel at 37° C, and could be optimized for IVD regeneration. The basic objective of this research is to characterize these new formulations and to determine those that present similar mechanical properties to human NP, while allowing the survival of encapsulated NP cells, and the synthesis of a tissue with a similar composition to that of native NP.

Materials and Methods

The hydrogels were prepared by mixing an acidic solution of chitosan and gelling agent solutions consisting of sodium hydrogen carbonate (SHC), a phosphate buffer (PB) and/or β -glycerophosphate (BGP). The composition of the four tested formulations is shown in Table 1.

Table 1 Composition of chitosan hydrogel formulations and their osmolality

Hydrogel	Chitosan	Gelling agent		ent	Osmolality (mOsmol/L)
	(w/v%)	composition		on	
		(M)			
		BGP	SHC	PB	
Formulation 1 (conventional formulation)	2	0.4		721	837 ± 12
Formulation 2	2	0.1	0.075		434 ± 25
Formulation 3	2		0.075	0.02	262 ± 3
Formulation 4	2		0.075	0.04	313 ± 4

Rheological tests with a rotational rheometer (Anton Paar) at room temperature (22° C) and body temperature (37° C) were performed to test the stability of the formulations at 22° C and the gelation profile at 37° C. Mechanical tests in unconfined compression on hydrogels incubated for 24 h at 37° C were performed to determine Young's secant modulus of each sample. These mechanical properties were compared to those of human NP in unconfined compression using an incremental stress relaxation test [6].

The osmolarity of these hydrogels was also measured. Then, bovine NP cells were encapsulated in these hydrogels and cultured for fourteen days at 37° C. Viability tests using AlamarBlue® and LIVE/DEAD® assay were performed to evaluate the metabolic activity of the encapsulated cells and their viability, respectively. The synthetic activity of these cells was also evaluated by measuring the amount of glycosaminoglycans (GAG) produced by the 1-dimethyl methylene (DMMB) assay. The injectability of the hydrogels was tested in explanted human discs using 25-gauge needles.



Results

All the hydrogel formulations, except for the conventional formulation (1), have a low storage modulus G' and are stable at 22° C. This modulus evolves rapidly at 37° C, confirming the thermosensitivity of the materials. The new hydrogel formulations have higher mechanical properties in compression than the conventional formulation (Figure 2 A, B, C). Stress relaxation tests show that two of these new formulations (2 and 3) present mechanical properties similar to human NP mechanical properties (FIG. 2 D).

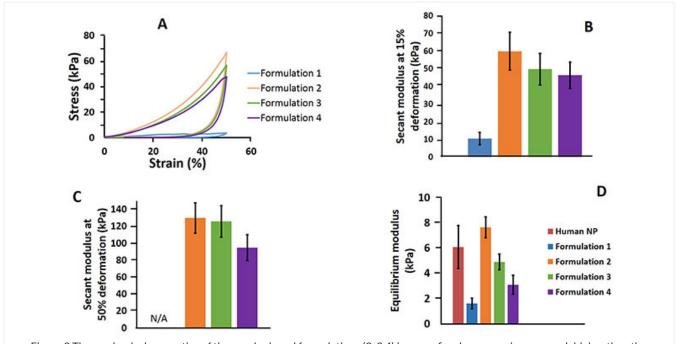


Figure 2 The mechanical properties of the new hydrogel formulations (2, 3.4) in unconfined compression are much higher than the conventional formulation (1). A) Stress strain curve, B) and C) Secant modulus at 15 and 50% D) Mechanical properties of hydrogel formulations compared to human nucleus pulposus in stress relaxation.

With regard to the biocompatibility results, the conventional formulation (1) is hypertonic with osmolality > 800 mOsmol/kg, while all the other formulations have osmolality values close to the physiological values within the disc (430 mOsmol/kg) [7]. A viability rate of approximately 85% of the encapsulated cells was observed, the highest metabolic activities being after 14 days of culture for formulations 2 and 3 (Figure 3). On the other hand, formulation 2 has the highest GAG production (Figure 4).

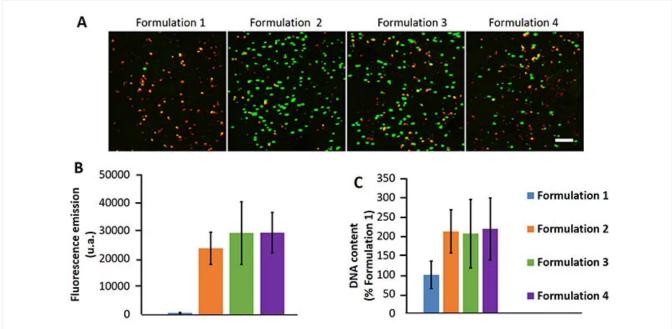


Figure 3 The new formulations show better biocompatibility after 14 days in culture A) Cell viability evaluated by LIVE/DEAD (red = dead cell, green = living cell) B) Metabolic activity of cells using AlamarBlue C) Amount of DNA present after 14 days

Hydrogel	Total quantity (μg)	Released quantity (µg)	Accumulated quantity in the hydrogel (µg)
Formulation 1	10.7 ± 1.2	5.7 ± 0.5	5.2 ± 0.7
Formulation 2	33.1 ± 0.9	12.1 ± 1.2	21.0 ± 1.1
Formulation 3	23.8 ± 1.9	4.1 ± 0.4	19.6 ± 1.6
Formulation 4	15.2 ± 1.1	0.8 ± 0.2	14.3 ± 1.2

Table 2 GAG production as per the formulation

Finally, injectability tests in explanted human discs show good retention of formulation 3. Not only does the initial viscosity of this gel allow injection with a smaller-diameter needle in addition to filling the defects present in the disc, but also shows gelation rapid enough to prevent leakage from the injection site (Figure 4).

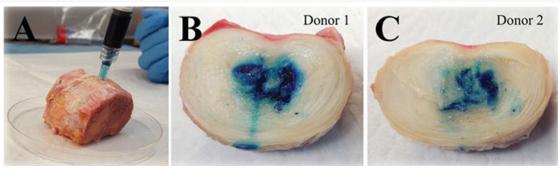


Figure 4 (A) Injection of the hydrogel (tinted with toluidine blue) into the NP of a human disc through the fibrous ring with a 25-gauge needle. The sample is kept at 37° C for 30 minutes. (B) 0.5 ml of hydrogel injected into disc 1. (C) 0.3 ml in disc 2.

Conclusion

This study concludes that Formulation 2 is a promising injectable matrix for the intervertebral disc repair because it exhibits mechanical properties similar to human NP and provides an adequate environment for cell survival as well as for the synthesis and retention of glycosaminoglycans, an important component of NP tissue. The remainder of the project will focus on studying the benefits of adding a bioactive molecule to cell synthesis activity and therefore to the regenerative capacity of the technology. Ultimately, this project will develop injectable hydrogels combining cells and bioactive agents for regenerative medicine, specifically intervertebral disc regeneration.

Additional Information

For more information on this research, please refer to the following paper:

Alinejad, Yasaman, Atma Adoungotchodo, Michael P Grant, Laura M Epure, John Antoniou, Fackson Mwale and Sophie Lerouge. 2018. "Injectable Chitosan Hydrogels with Enhanced Mechanical Properties for Nucleus Pulposus Regeneration." Tissue Engineering Part A. doi: 10.1089/ten.tea.2018.0170



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